

16<sup>th</sup>  
**CONGRESS**  
*Lung* **ON**  
**CANCER**

BARCELONA  
27 / 28  
NOVEMBER 2025

# Pitfalls of liquid biopsy

**Atocha Romero**

*Hospital Universitario Puerta de Hierro*

# CONFLICTO DE INTERESES

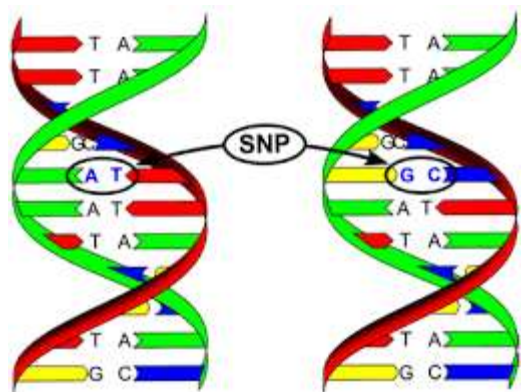
- Grants from J&J and EMQN
- Consultant or advisory roles for Menarini and J&J
- Presentation for Illumina, Health in Code, and ThermoFisher Scientific, Menarini
- Support for attending meetings and/or travel from ThermoFisher Scientific, Bristol Myers Squibb, and Takeda

# Pitfalls of liquid biopsy



- 1. Sensitivity and Detection Limits**
- 2. Specificity and False Positives**
- 3. Tumor Fraction estimation**
- 4. Pre-analytical and Analytical Variability**
- 5. Limited Clinical Validation and Utility:**

# Sensitivity and Detection Limits. Fusion detection



## Indel examples

wild-type sequence

ATCTTCAGCCATAAAAGATGAAGTT

3 bp deletion

ATCTTCAGCCCAAAGATGAAGTT

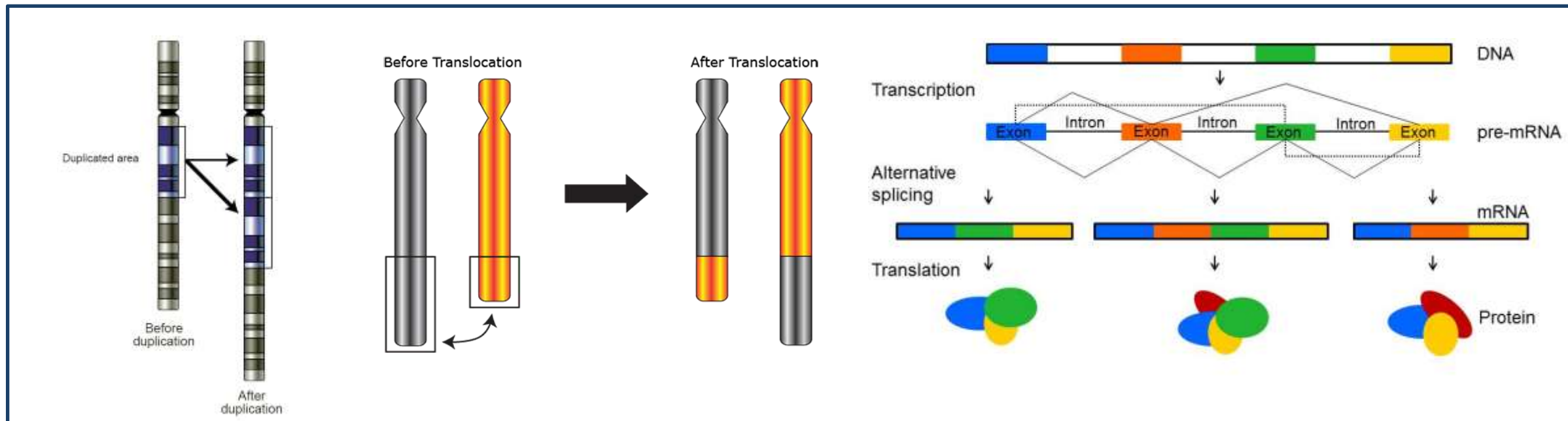
4 bp insertion (orange)

ATCTTCAGCCCATATGTGAAAGATGAAGTT

Normal: See the dog run.

Deletion: ~~S~~et hed ogr un.

Insertion: See eth edo gru n.



# Sensitivity and Detection Limits. Fusion detection

## Sensitivity of next-generation sequencing assays detecting oncogenic fusions in plasma cell-free DNA



Julianna G. Supplee<sup>a</sup>, Marina S.D. Milan<sup>b</sup>, Lee P. Lim<sup>c</sup>, Kristy T. Potts<sup>c</sup>, Lynette M. Sholl<sup>d</sup>, Geoffrey R. Oxnard<sup>b</sup>, Cloud P. Paweletz<sup>a,\*</sup>

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<sup>b</sup> Lowe Center for Thoracic Oncology, Dana-Farber Cancer Institute, 450 Brookline Avenue, Boston, MA, 02215, USA

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### ARTICLE INFO

#### Keywords:

NSCLC  
Cell-free DNA  
Next-generation sequencing  
Gene fusions

### ABSTRACT

**Objectives:** Plasma genotyping represents an opportunity for convenient detection of clinically actionable mutations in advanced cancer patients, such as has been well-documented in non-small cell lung cancer (NSCLC). Oncogenic gene fusions are complex variants that may be more challenging to detect by next-generation sequencing (NGS) of plasma cell-free DNA (cfDNA). Rigorous evaluation of plasma NGS assays in the detection of fusions is needed to maximize clinical utility.

**Materials and methods:** Additional plasma was collected from patients with advanced NSCLC and *ALK*, *ROS1*, or *RET* gene fusions in tissue who had undergone clinical plasma NGS using Guardant360™ (G360, Guardant Health). We then sequenced extracted cfDNA with a plasma NGS kit focused on known driver mutations in NSCLC (ctDx-Lung, Resolution Bioscience) with cloud-based bioinformatic analysis and blinded variant calling.

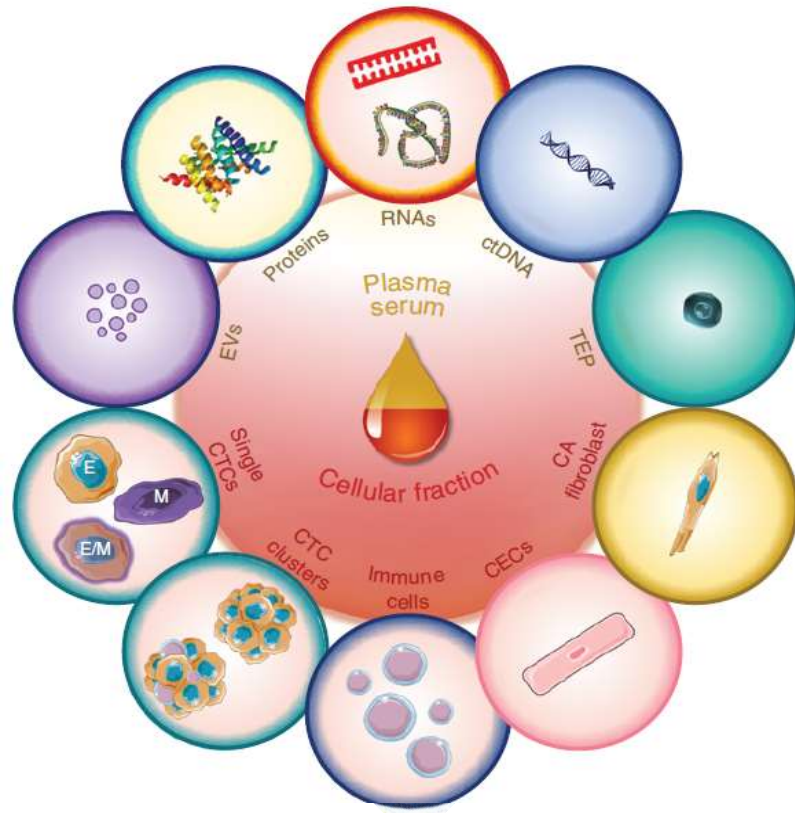
**Results:** Of 16 patients assayed known to harbor an *ALK*, *ROS1*, or *RET* in tumor, G360 detected fusions in 7 cases, ctDx-Lung detected fusions in 13 cases, and 3 cases were detected by neither. Of the 7 fusions detected by both assays, G360 reported lower mutant allelic fractions (AF). In cases missed by G360, tumor derived *TP53* mutations were often detected confirming presence of tumor DNA. Raw sequencing data showed that inverted or out-of-frame variants were overrepresented in cases detected using ctDx-Lung but not by G360.

**Conclusion:** Focusing on complex, clinically actionable mutations using tumor as a reference standard allows for evaluation of technical differences in plasma NGS assays that may impact clinical performance. Noting the heterogeneity of fusion sequences observed in NSCLC, we hypothesize that differences in hybrid capture techniques and bioinformatic calling may be sources of variations in sensitivity among these assays.

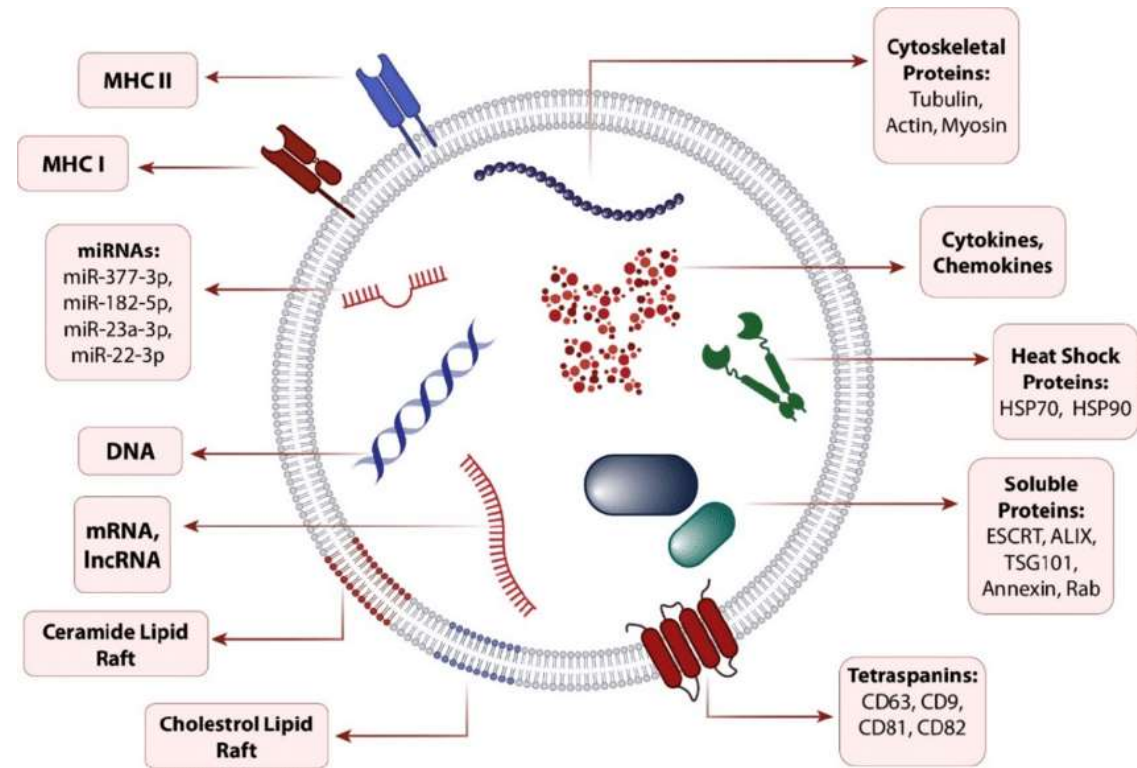


19%

# Sensitivity and Detection Limits. Fusion detection. EVs



Alix-Panabières, C et al. Cancer Discov. 2021;11:858-873





Int J Mol Sci. 2022 Jun 3;23(11):6273. doi: 10.3390/ijms23116273.

# EVs

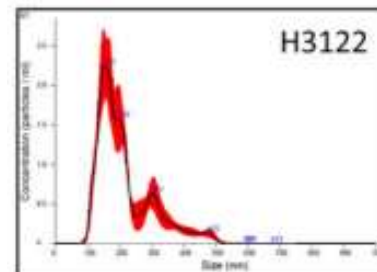
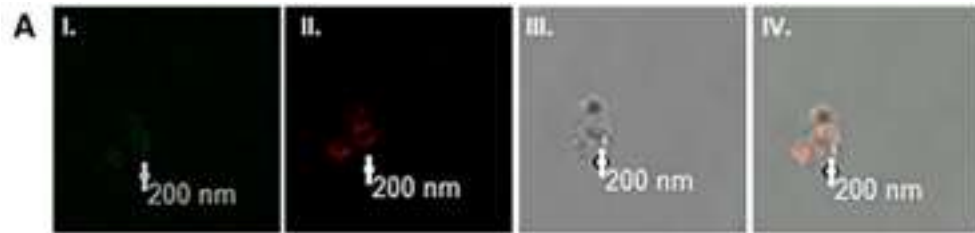
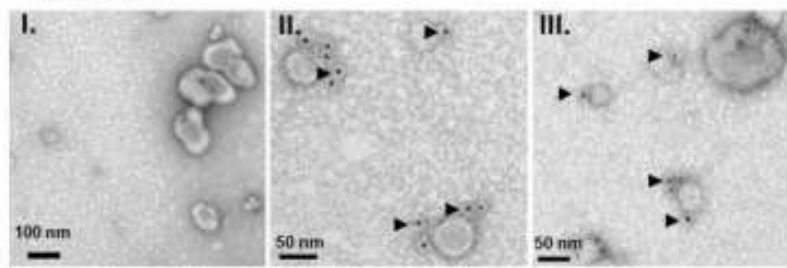
Clinical Chemistry 68:6  
1-12 (2022)

Molecular Diagnostics and Genetics

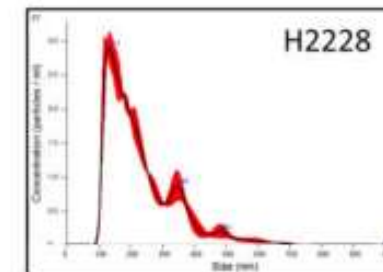
## ALK-Fusion Transcripts Can Be Detected in Extracellular Vesicles (EVs) from Nonsmall Cell Lung Cancer Cell Lines and Patient Plasma: Toward EV-Based Noninvasive Testing

Estela Sánchez-Herrero,<sup>a,b,†</sup> Carmen Campos-Silva,<sup>c,†</sup> Yaiza Cáceres-Martell,<sup>c</sup> Lucía Robado de Lope,<sup>a</sup> Sandra Sanz-Moreno,<sup>a</sup> Roberto Serna-Blasco,<sup>a</sup> Alejandro Rodríguez-Festa,<sup>a</sup> Dunixie Ares Trotta,<sup>a</sup> Paloma Martín-Acosta,<sup>d</sup> Cristina Patiño,<sup>a</sup> María José Coronado,<sup>f</sup> Alexandra Beneitez,<sup>g</sup> Ricardo Jara,<sup>g</sup> Nerea Lago-Baameiro,<sup>h</sup> Tamara Camino,<sup>h</sup> Alberto Cruz-Bermúdez,<sup>a</sup> María Pardo,<sup>h</sup> Víctor González-Rumayor,<sup>b</sup> Mar Valés-Gómez ,<sup>c,\*</sup> Mariano Provencio,<sup>a,j</sup> and Atocha Romero <sup>a,i,\*</sup>

### B TEM



Mean $\phi$	215.9 +/- 3.4 nm
Mode $\phi$	154.7 +/- 6.3 nm
Particles/ $\mu$ L	$4.6 \cdot 10^8$ +/- $1.14 \cdot 10^8$



Mean $\phi$	228.8 +/- 4.9 nm
Mode $\phi$	126.6 +/- 4.8 nm
Particles/ $\mu$ L	$3.2 \cdot 10^8$ +/- $7.85 \cdot 10^7$

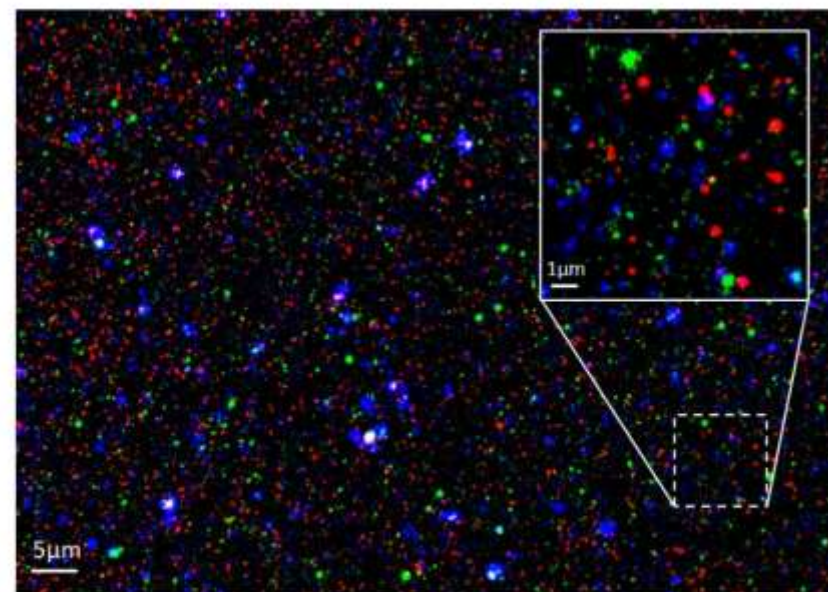
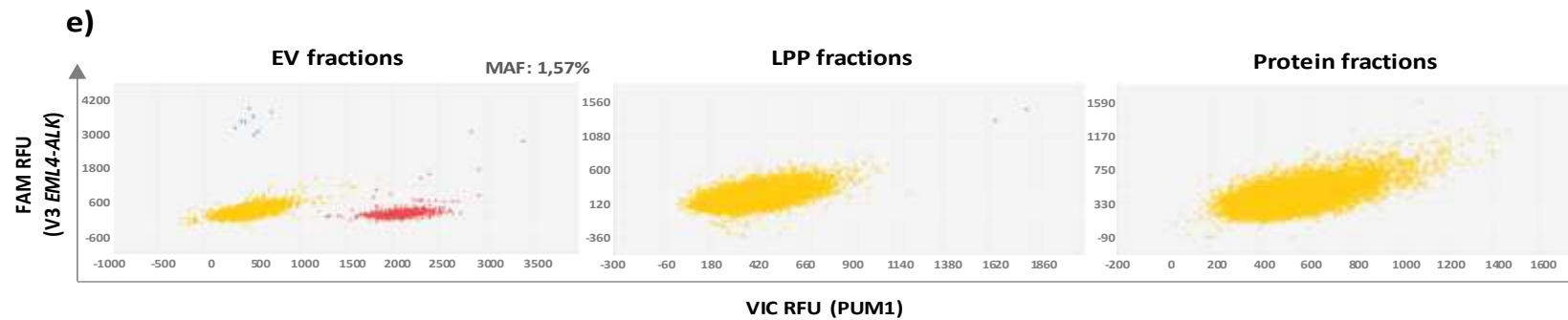
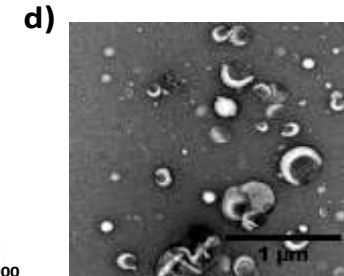
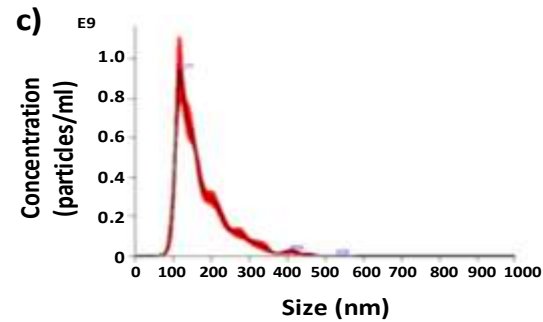
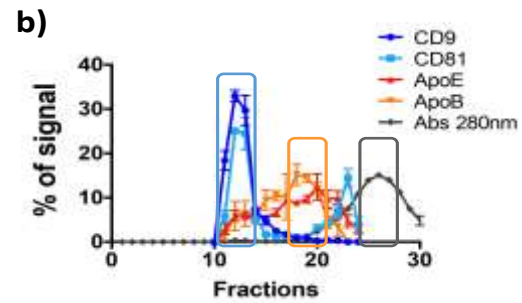
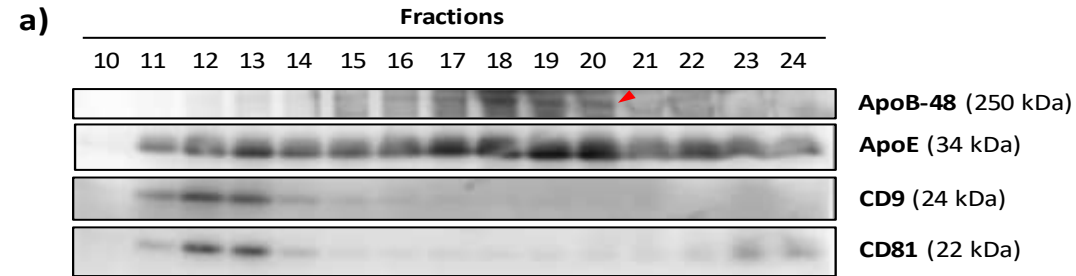
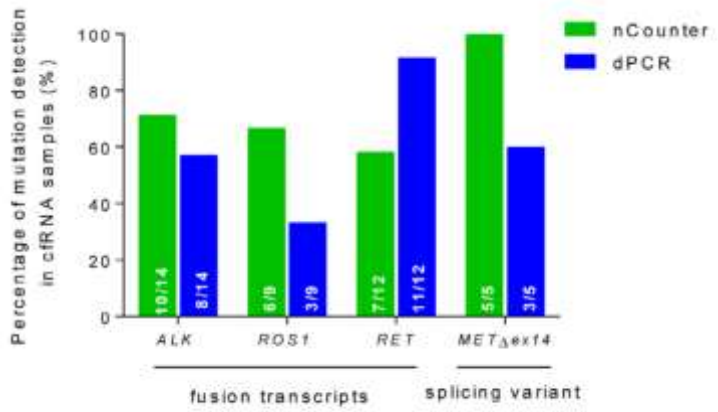
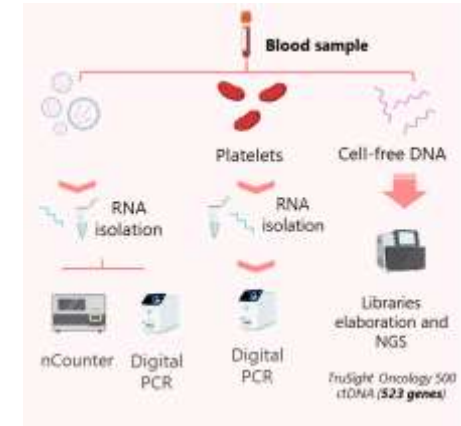
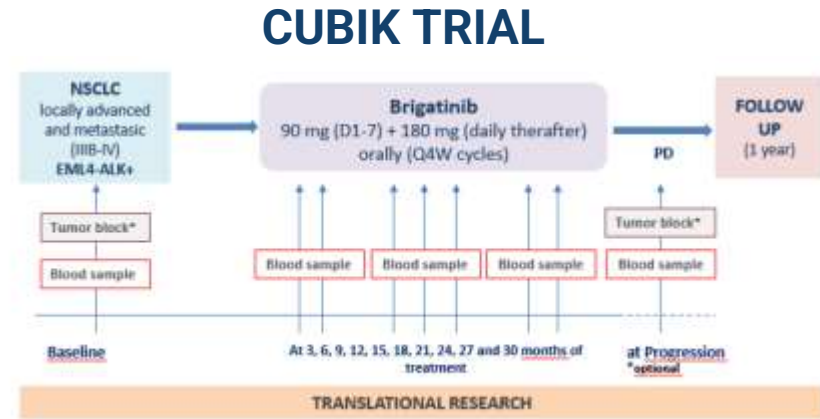
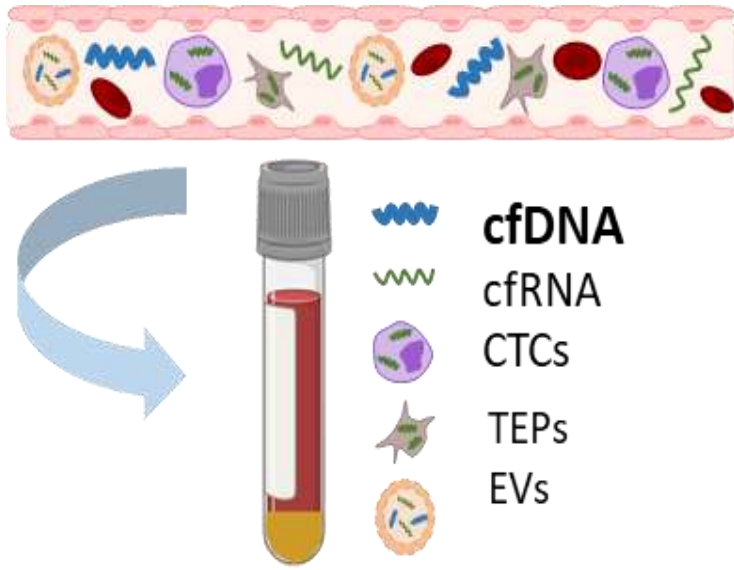


Figure S2. Characterization of EV-enriched preparations from NSCLC plasma samples by ExoView platform. CD9-captured EVs image in fluorescence mode: CD63 in red, CD9 in blue, ALK\* in green, CD63-ALK\* in yellow, CD63-CD9 in purple, ALK\*-CD9 in light blue, and CD63-CD9-ALK\* in white. ALK\*, ALK fusion protein.

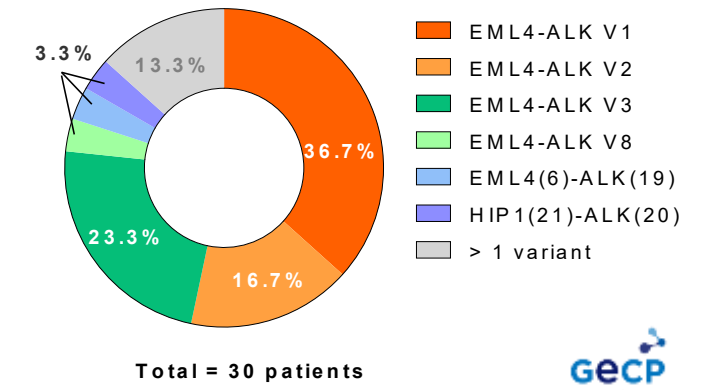
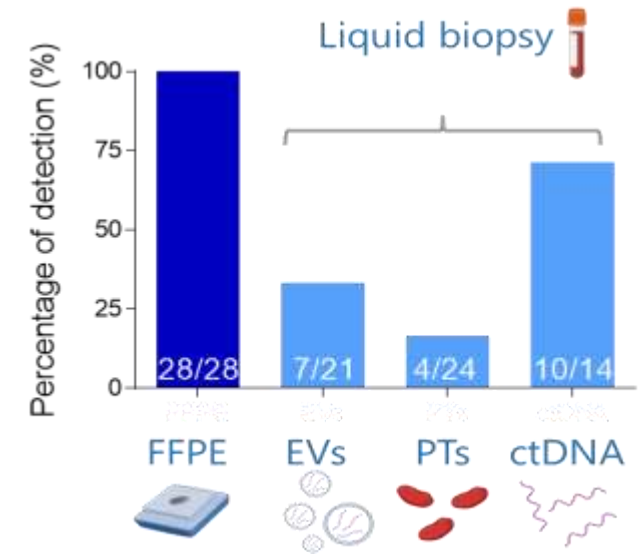
# EVs



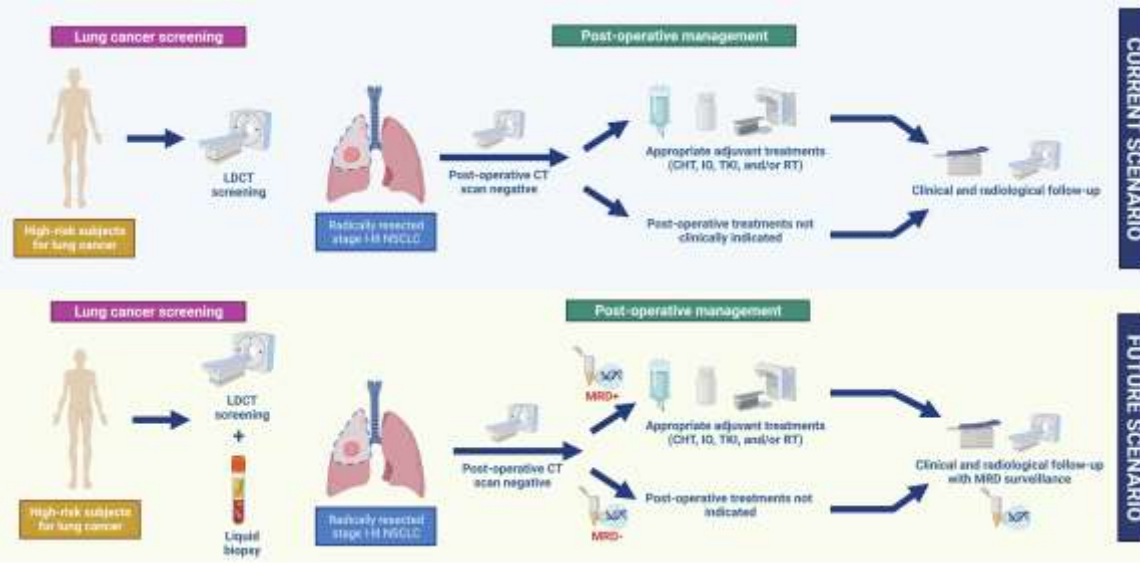
# cfRNA, Evs and platelets



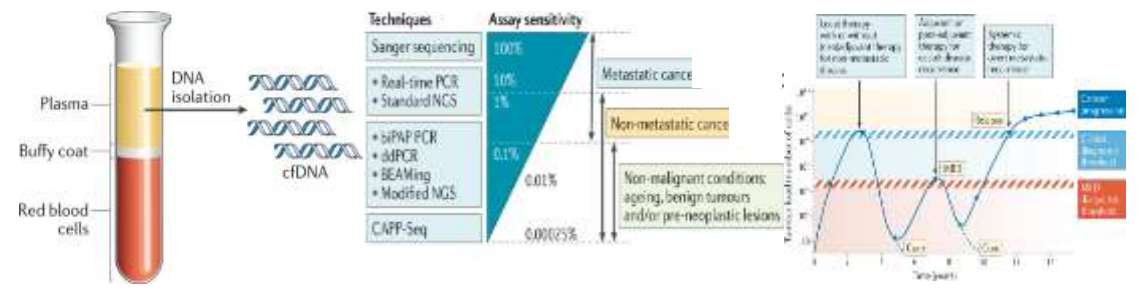
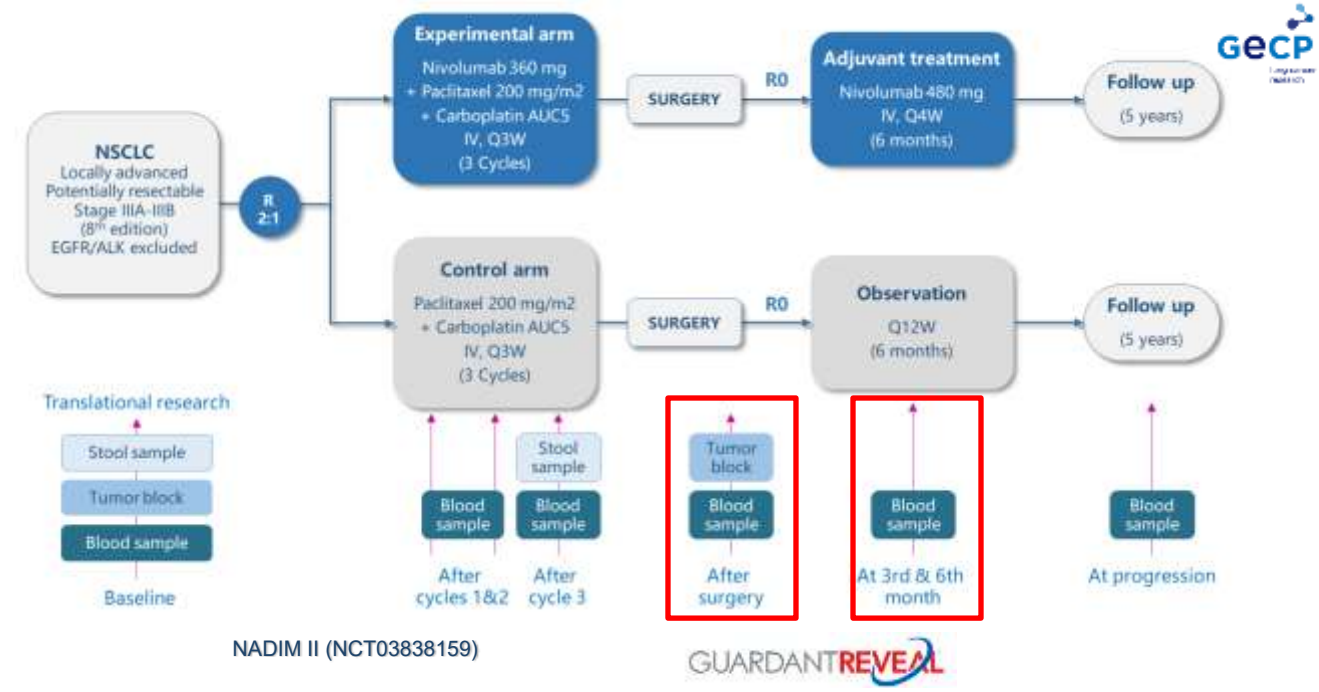
<https://pubmed.ncbi.nlm.nih.gov/37243883/>



# Sensitivity and Detection Limits MRD in NSCLC



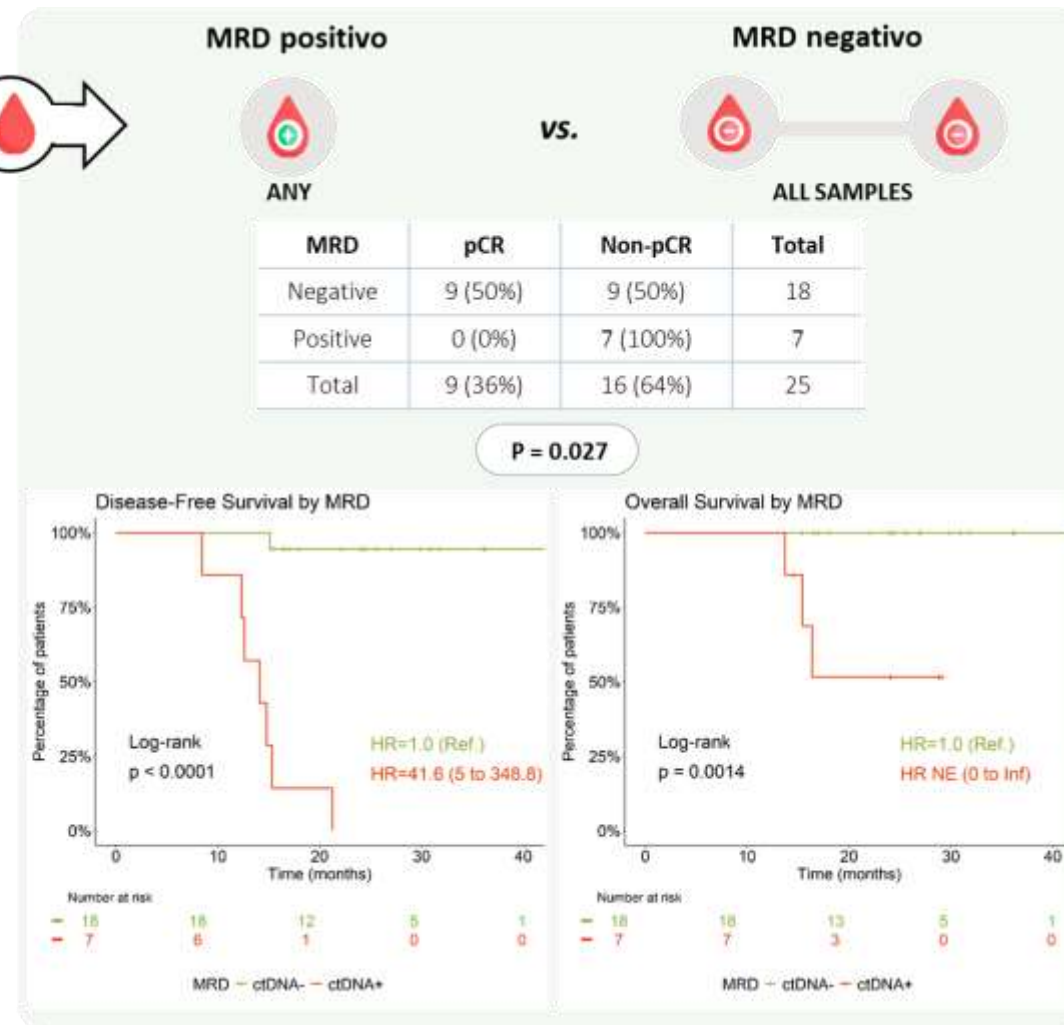
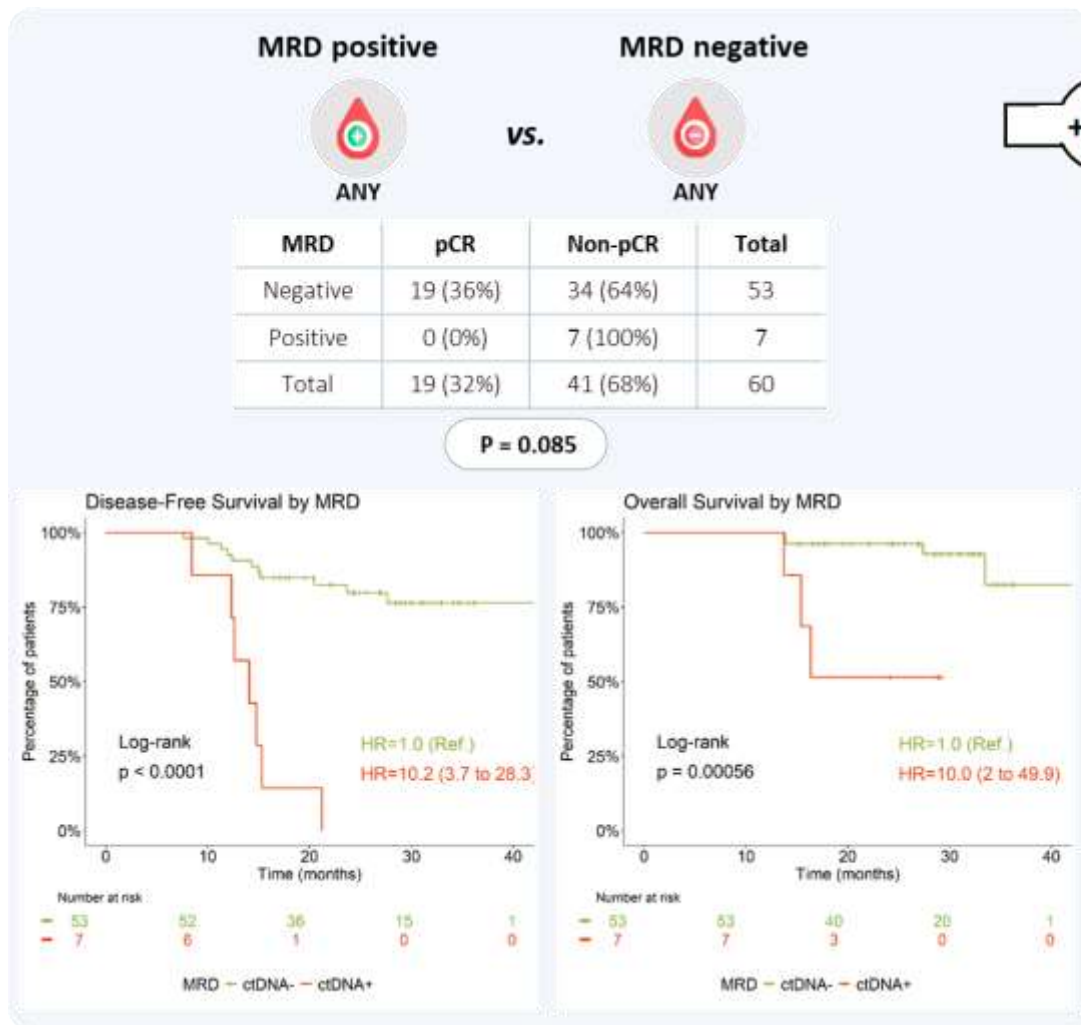
<https://www.thelancet.com/journals/eblm/article/PIIS2352-3964%2823%2900170-6/fulltext>



Pantel, K et al. Nat Rev Clin Oncol. 2019;16:409-424.

10% detection rate

# MRD in NADIMII



# MRD in NADIMII

MRD positive

MRD negative



vs.

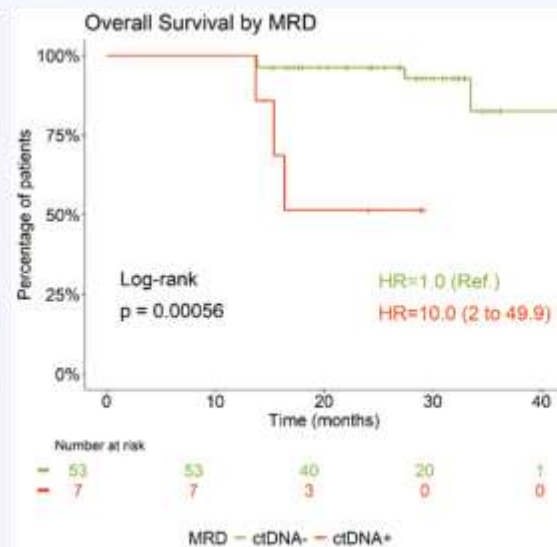
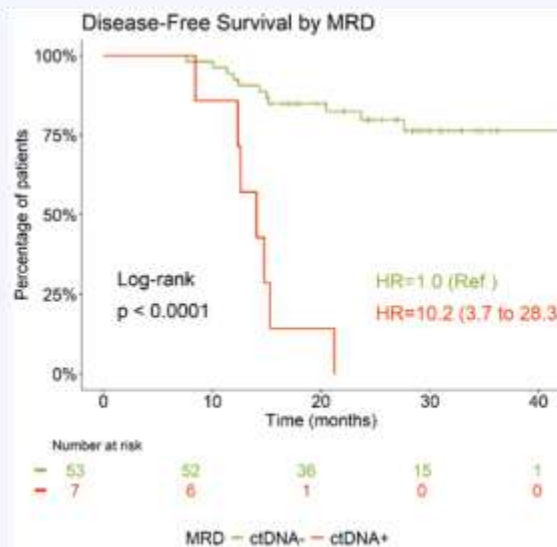


ANY

ANY

MRD	pCR	Non-pCR	Total
Negative	19 (36%)	34 (64%)	53
Positive	0 (0%)	7 (100%)	7
Total	19 (32%)	41 (68%)	60

P = 0.085



MRD positivo

MRD negativo



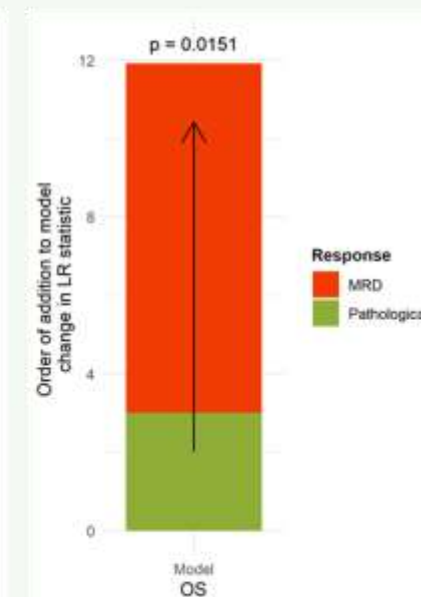
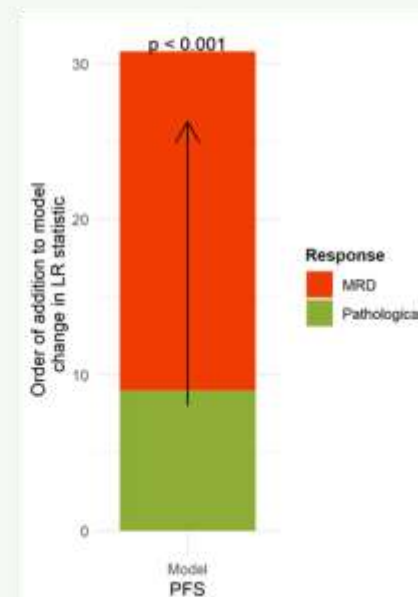
vs.



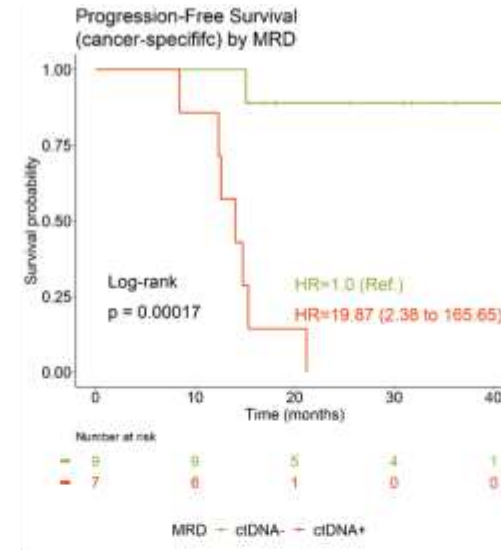
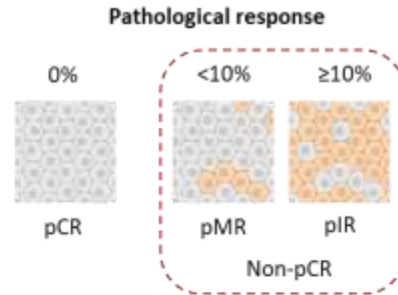
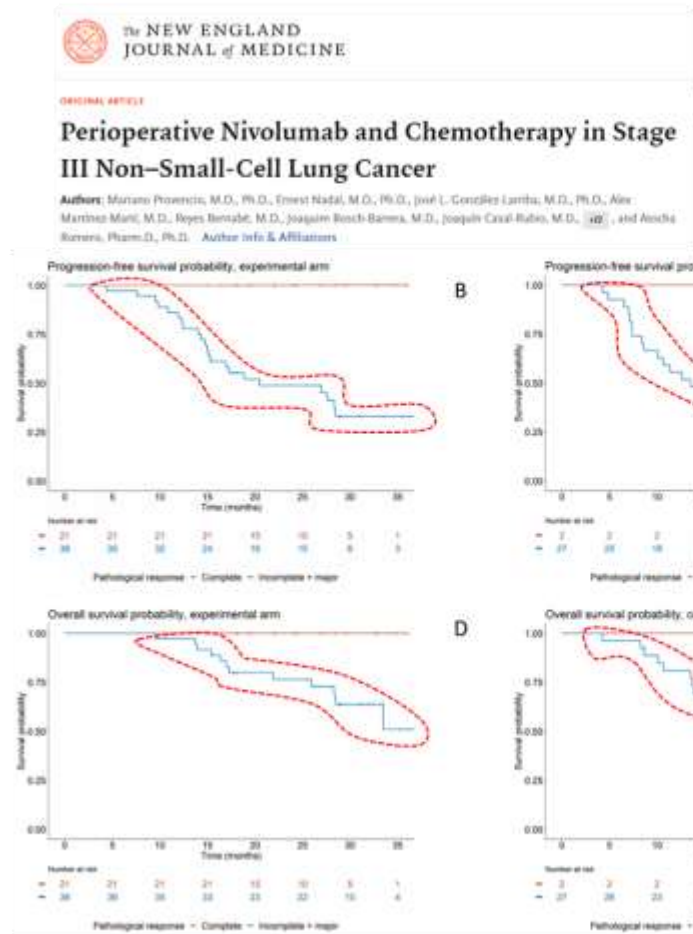
ANY

ALL SAMPLES

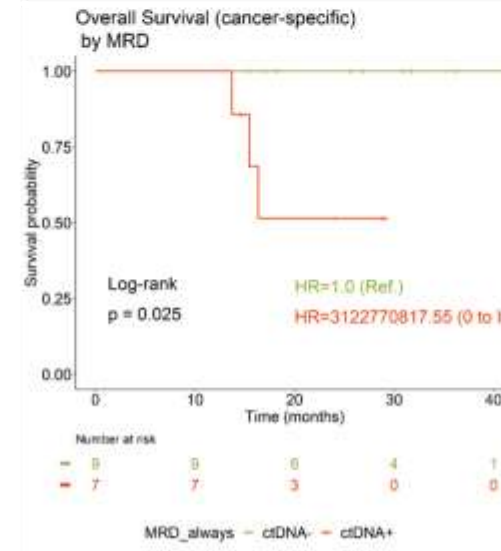
**Does MRD detection significantly improve pathological response prognosis?**



# MRD in NADIMII



← Non-pCR patients with good prognosis



← Non-pCR patients with good prognosis

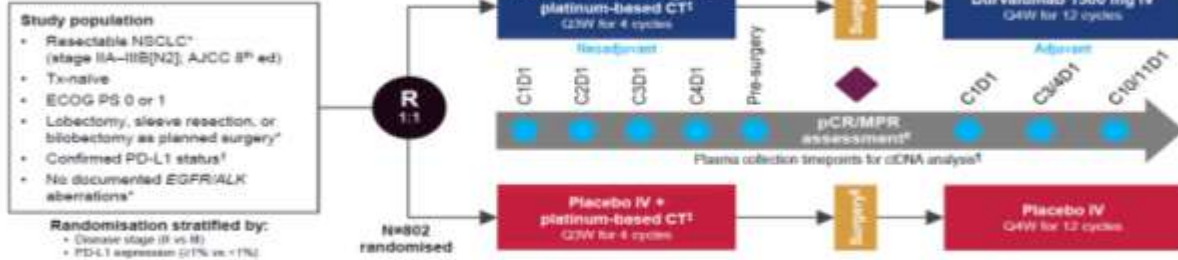
M Provencio et al., NEJM 2023 (PMID: 37379158)

**PRE-PUBLICATION DATA – DO NOT CITE OR SHARE.** Unauthorized distribution is prohibited.

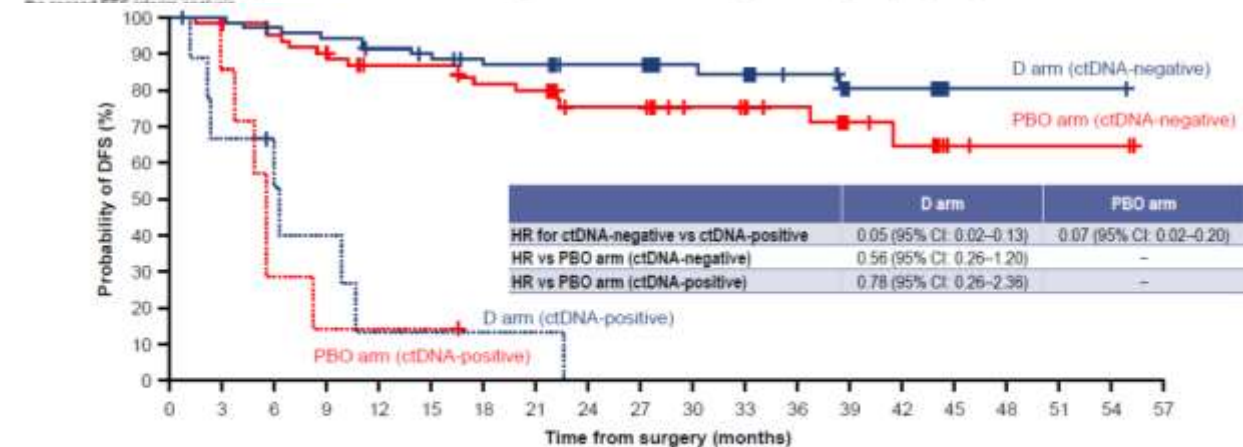
# MRD in NSCLC

## AEGEAN Study Design

Phase 3, global, randomised, double-blind, placebo-controlled study



- Plasma samples were collected at protocol-specified timepoints, including prior to each neoadjuvant Tx cycle, surgery, and adjuvant Tx at select cycles
- ctDNA analysis was performed using Invitae Personalized Cancer Monitoring™, a tumour-informed MRD assay,<sup>11</sup> with the exploratory analyses reported here based on data from



	D arm	PBO arm
HR for ctDNA-negative vs ctDNA-positive	0.05 (95% CI: 0.02-0.13)	0.07 (95% CI: 0.02-0.20)
HR vs PBO arm (ctDNA-negative)	0.56 (95% CI: 0.26-1.20)	-
HR vs PBO arm (ctDNA-positive)	0.78 (95% CI: 0.26-2.38)	-

No. at risk	72	71	68	66	63	61	58	56	48	48	32	30	24	14	1	1	1	1	0
D arm (ctDNA-negative)	72	6	5	3	1	1	1	1	0	0	0	0	0	0	0	0	0	0	0
D arm (ctDNA-positive)	9	6	5	3	1	1	1	1	0	0	0	0	0	0	0	0	0	0	0
PBO arm (ctDNA-negative)	62	61	58	55	50	50	46	45	32	32	25	24	18	12	10	3	2	2	2
PBO arm (ctDNA-positive)	7	6	2	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0

Associations of ctDNA Clearance during Neoadjuvant Treatment with Pathological Response and Event-free Survival in Patients with Resectable NSCLC. Martin Reck. ESMO 2024

nature medicine



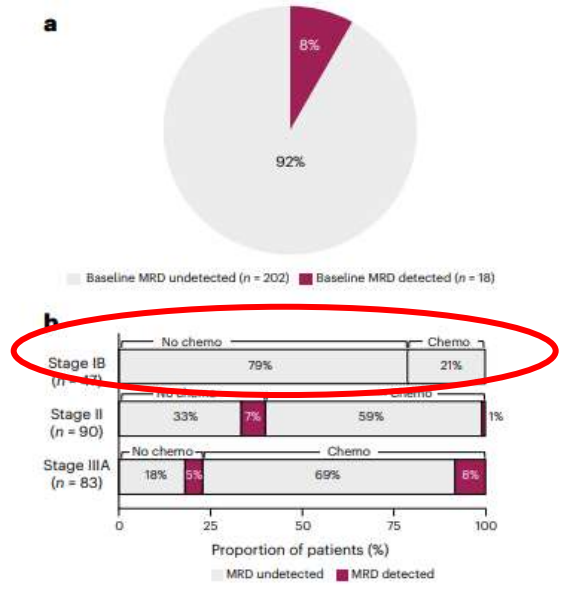
Article <https://doi.org/10.1038/s41591-025-03577-y>

## Molecular residual disease analysis of adjuvant osimertinib in resected EGFR-mutated stage IB-IIIa non-small-cell lung cancer

### MRD status at randomization:

- Detection rate: 8% (18/220)
- 4% receiving Osimertinib (5/12)
- 12% receiving placebo (13/108)

DFS status, n	Event (n=96)	MRD status, n	
		Detected (n=68)	Undetected (n=152)
Censor (n=124)	6	118	
Concordance of MRD with DFS, % (95% CI)			
PPA (sensitivity)	65 (55-74)		
NPA (specificity)	95 (91-99)		
PPV	81 (71-88)		
NPV	78 (71-84)		
OPA	68 (57-81)		



Stage	No chemo	Chemo
Stage IB (n = 47)	79%	21%
Stage II (n = 90)	33%	7%
Stage IIIA (n = 83)	18%	5%

# Use of Circulating Tumor DNA for Curative-Intent Solid Tumor Drug Development Guidance for Industry

*Additional copies are available from:*

Office of Communications, Division of Drug Information  
Center for Drug Evaluation and Research  
Food and Drug Administration  
10001 New Hampshire Ave., Hillandale Bldg., 4<sup>th</sup> Floor  
Silver Spring, MD 20993-0002  
Phone: 855-543-3784 or 301-796-3400; Fax: 301-431-6353  
Email: [druginfo@fda.hhs.gov](mailto:druginfo@fda.hhs.gov)  
<https://www.fda.gov/drugs/guidance-compliance-regulatory-information/guidance-drugs>

Office of Communication, Outreach and Development  
Center for Biologics Evaluation and Research  
Food and Drug Administration  
10903 New Hampshire Ave., Bldg. 71, Room 3128  
Silver Spring, MD 20993-0002  
Phone: 800-833-4709 or 240-402-8010  
Email: [ocou@fda.hhs.gov](mailto:ocou@fda.hhs.gov)  
<https://www.fda.gov/vaccines-blood-biologics/guidance-compliance-regulatory-information-biologics/biologics-guidance>

Office of Policy  
Center for Devices and Radiological Health  
Food and Drug Administration  
10903 New Hampshire Ave., Bldg. 66, Room 5431  
Silver Spring, MD 20993-0002  
Phone: 301-796-5900  
E-mail: [CDRH-Guidance@fda.hhs.gov](mailto:CDRH-Guidance@fda.hhs.gov)  
<http://www.fda.gov/MedicalDevices/DeviceRegulationandGuidance/Guidance/Document/default.htm>

U.S. Department of Health and Human Services  
Food and Drug Administration  
Oncology Center of Excellence (OCE)  
Center for Devices and Radiologic Health (CDRH)  
Center for Drug Evaluation and Research (CDER)  
Center for Biologics Evaluation and Research (CBER)  
November 2024

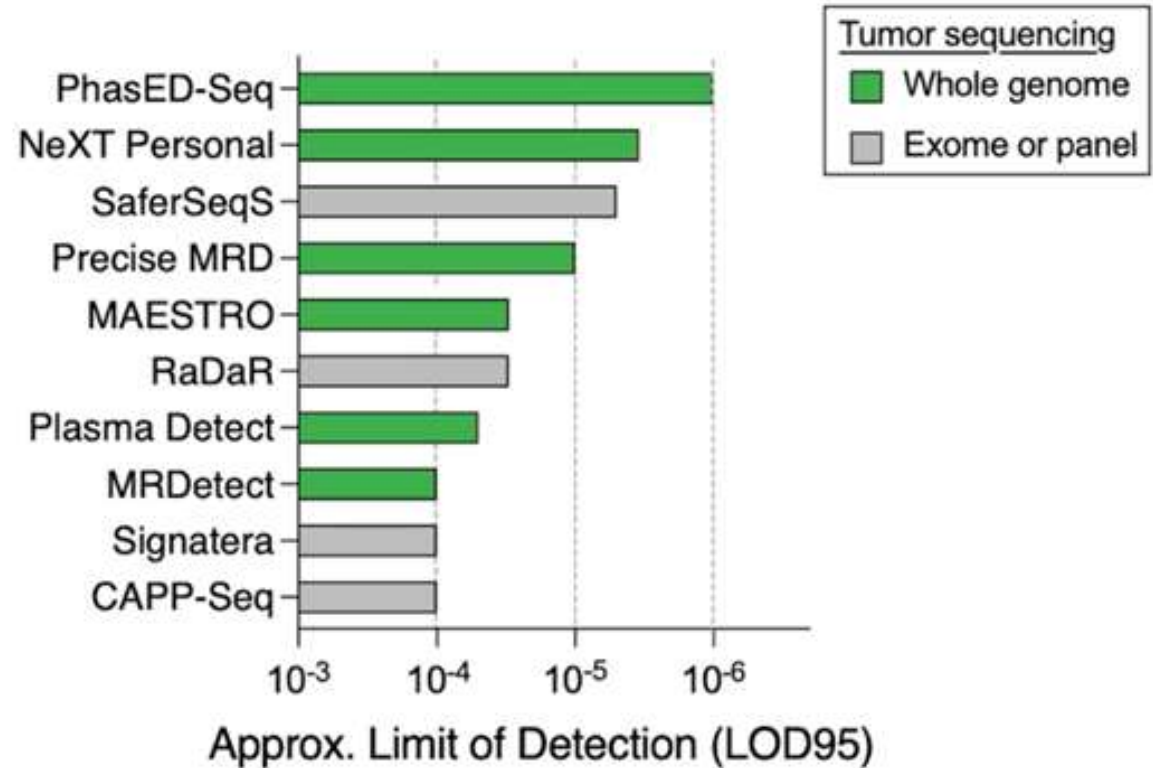
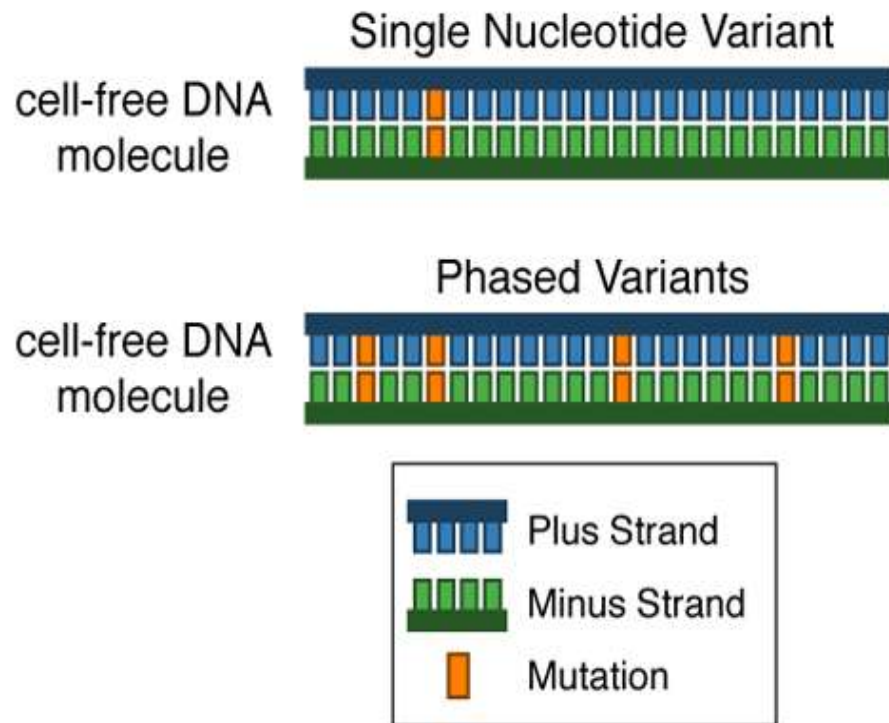
ctDNA MRD could be used for treatment optimization, to add on therapy for patients who are at higher risk of disease recurrence (i.e., ctDNA MRD positive) or to de-escalate therapy for patients with lower risk of disease recurrence (i.e., ctDNA MRD negative)

The MRD assay should have high sensitivity and negative predictive value (NPV) for supporting de-escalation of treatment and high specificity and positive predictive value (PPV) for supporting escalation of treatment.

- MRD detection with 1st generation assays has a high positive predictive value.
- Sensitivity remains a challenge

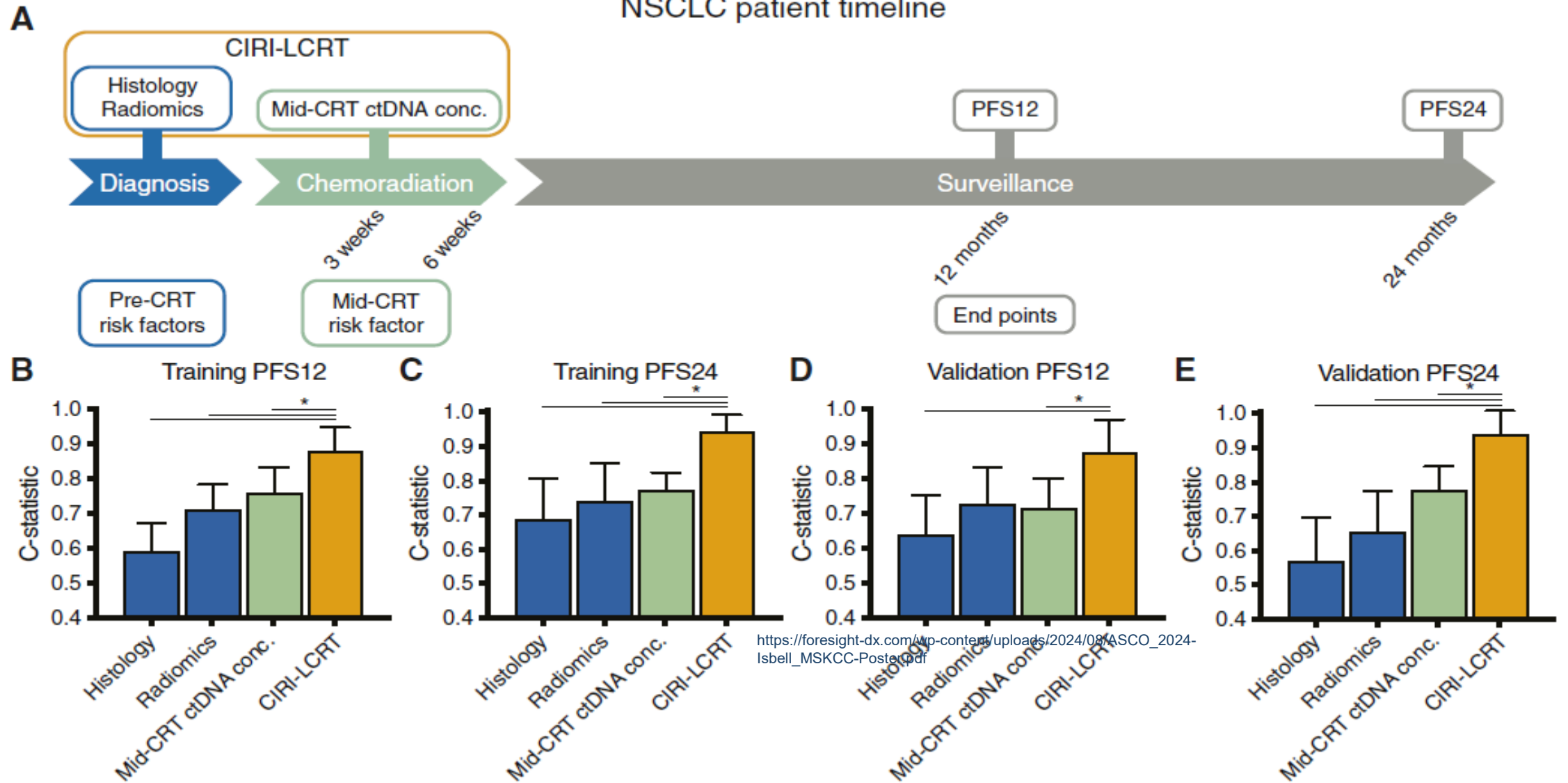
# Approaches to improve LOD

## Phased variants in ctDNA



[https://foresight-dx.com/wp-content/uploads/2024/08/ASCO\\_2024-Isbell\\_MSKCC-Poster.pdf](https://foresight-dx.com/wp-content/uploads/2024/08/ASCO_2024-Isbell_MSKCC-Poster.pdf)

# Approaches to improve LOD

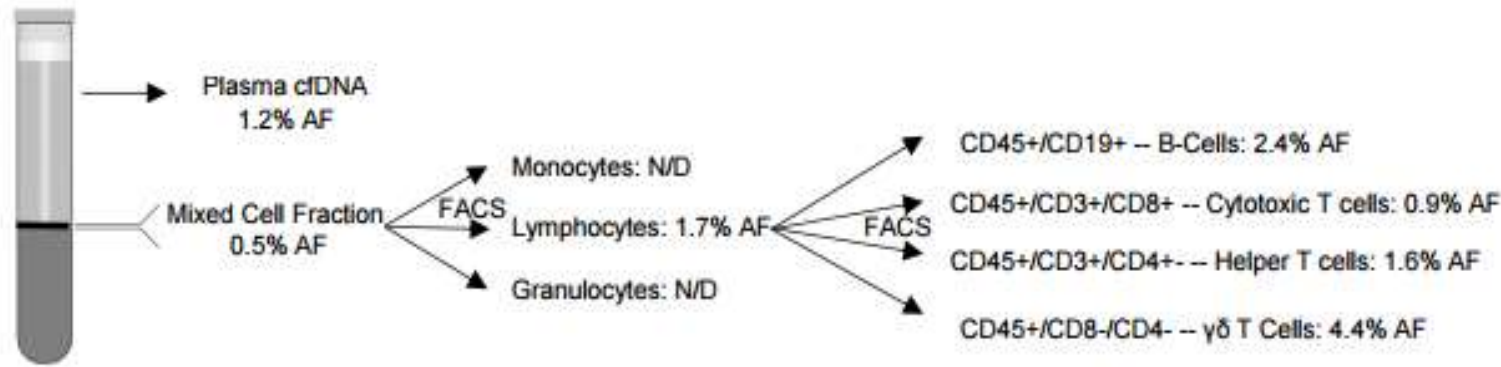


# Specificity and False Positives

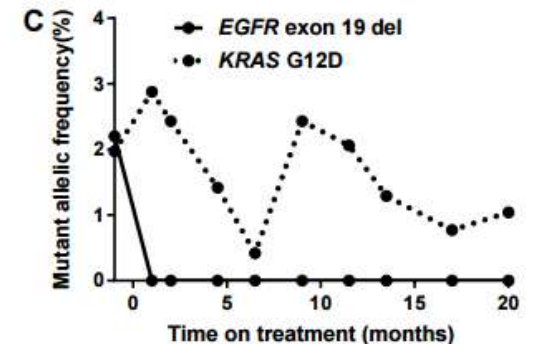
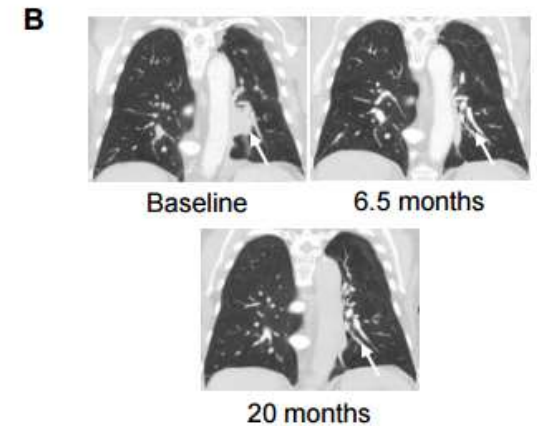
## Clonal haematopoiesis

- CHIP is defined by the presence of somatic mutation in blood or BM but without other diagnostic criteria for a haematological malignancy.
- More frequent in aged patients, patients with solid tumours
- More likely to be detected with deeper sequencing approaches.
- May account for false positive ctDNA sequencing results.

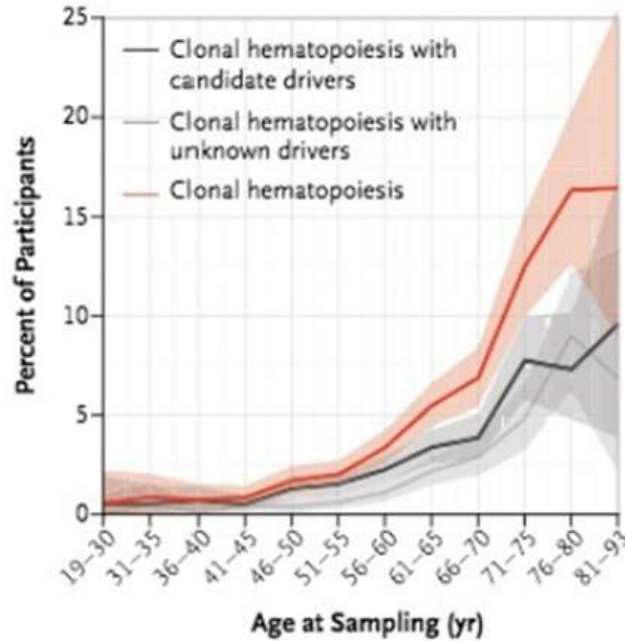
KRAS G12D



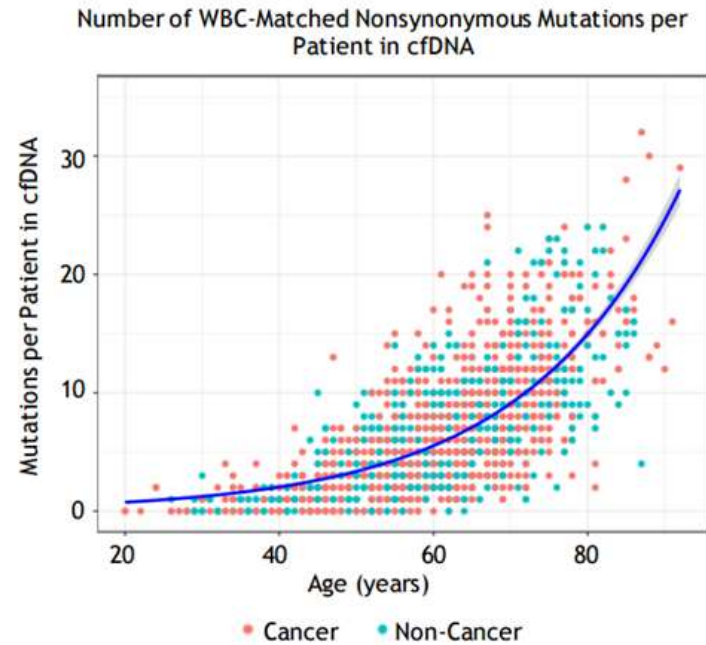
Hu Y et al. False-Positive Plasma Genotyping Due to Clonal Hematopoiesis. Clin Cancer Res. 2018 Mar 22.



# Specificity and False Positives

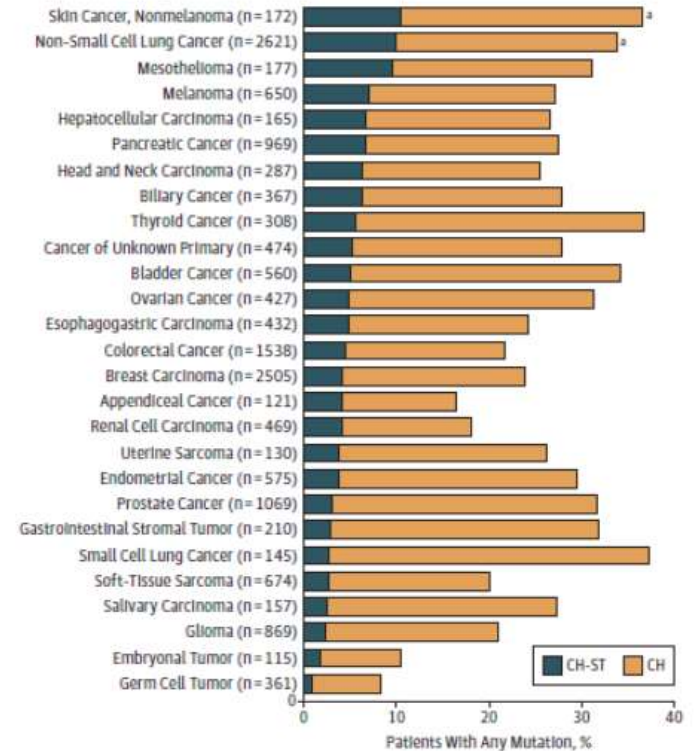


Genovese G., *et al.* 2014 NEJM



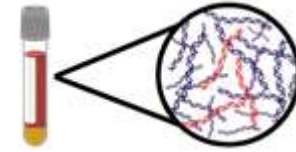
Charles Swanton *et al.* Journal of Clinical Oncology 36, no. 15\_suppl (May 20 2018) 12003-12003.

B Frequency by cancer type



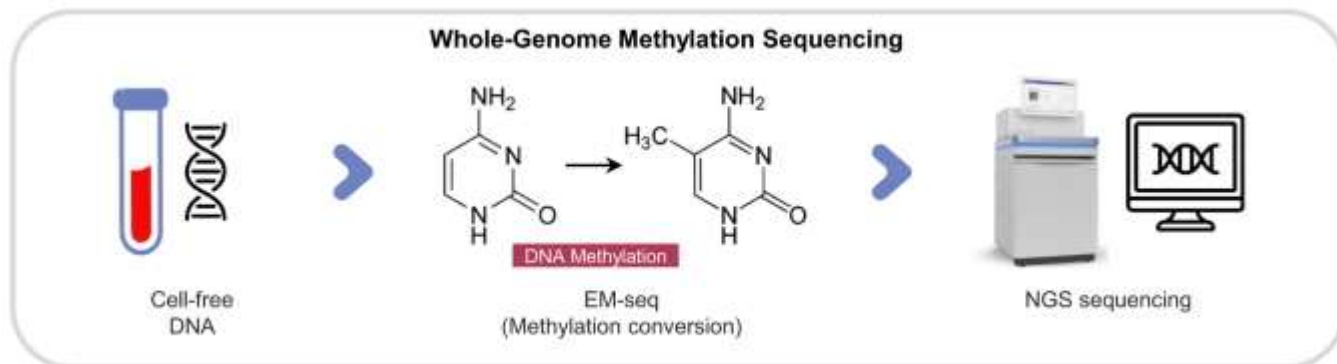
# Tumor Fraction estimation

Methods for estimating tumor fraction in circulating tumor DNA (ctDNA) using liquid biopsy

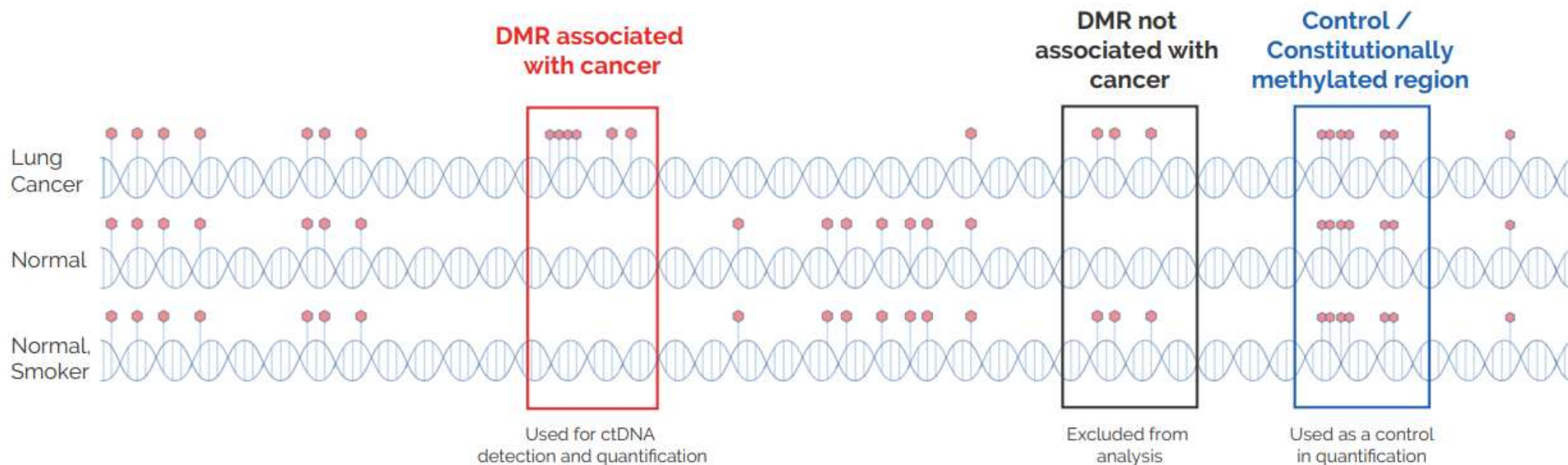


1. **Variant Allele Frequency (VAF) of Tumor-Specific Mutations**
2. **Genome-Wide Copy Number Alterations (CNAs):** The amplitude of these alterations in cfDNA is used to infer the fraction of ctDNA, especially in tumors with high chromosomal instability
3. **Methylation Profiling**
4. **Fragment Size Analysis**
5. **Bioinformatic Deconvolution**

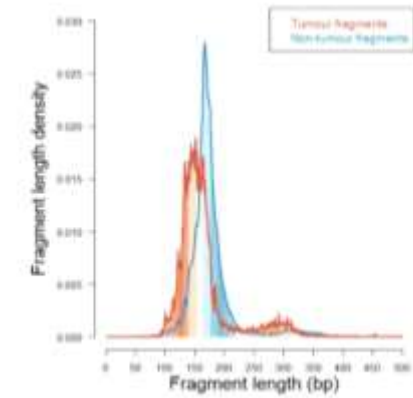
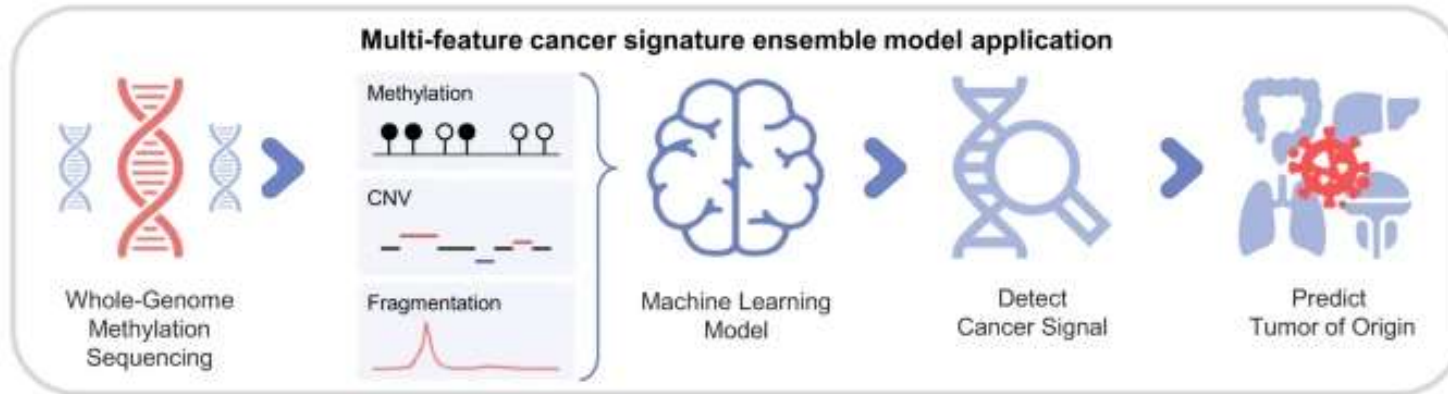
# TUMOR FRACTION. METILATION



<https://www.nature.com/articles/s12276-023-01119-5>

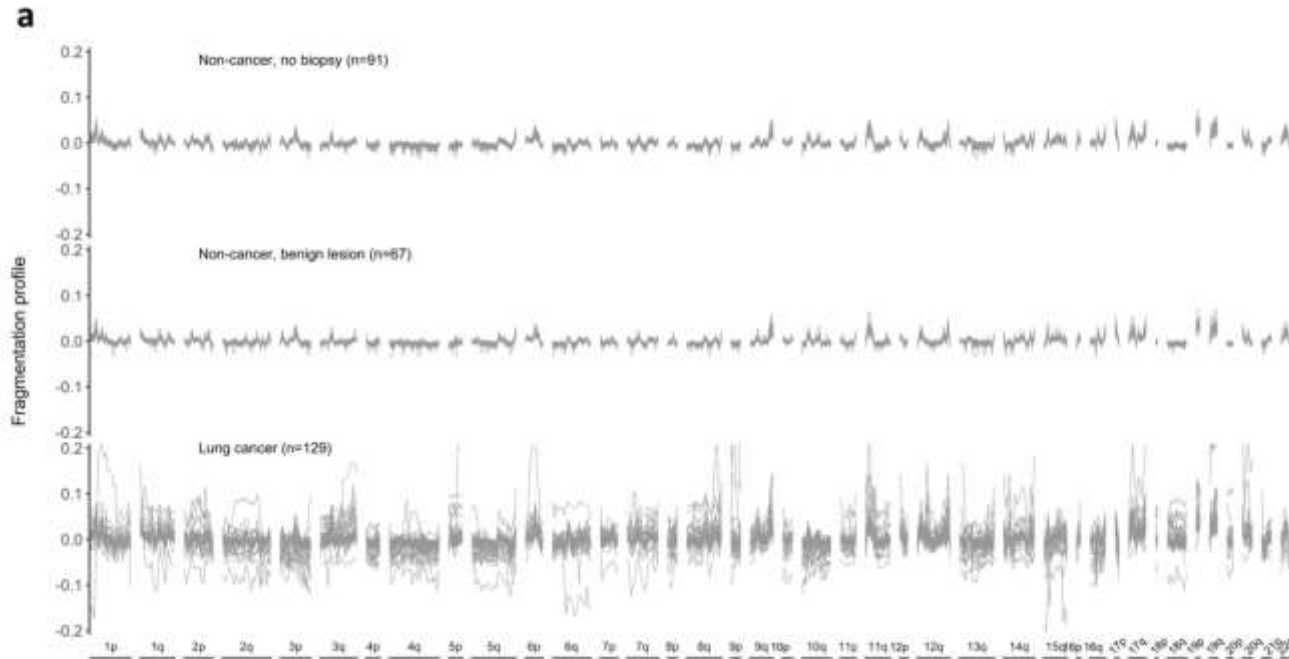


# FRAGMENTOMICS

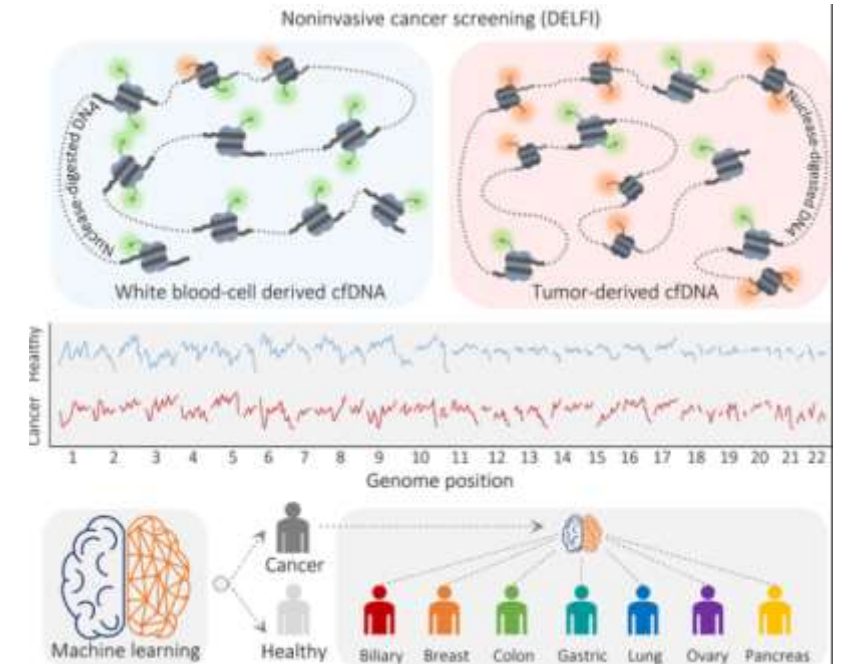


Vessies et al., *Molecular Oncology* 16:2719-2732 (2022)

<https://www.nature.com/articles/s12276-023-01119-5>

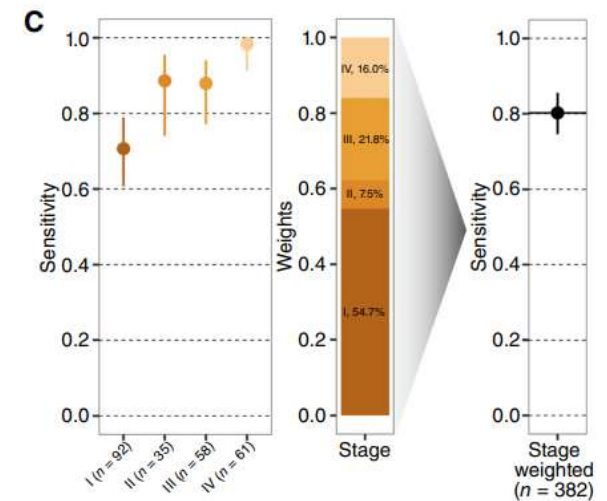
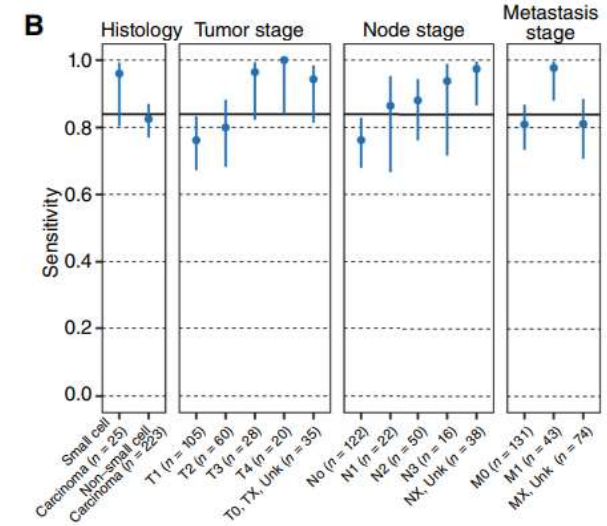
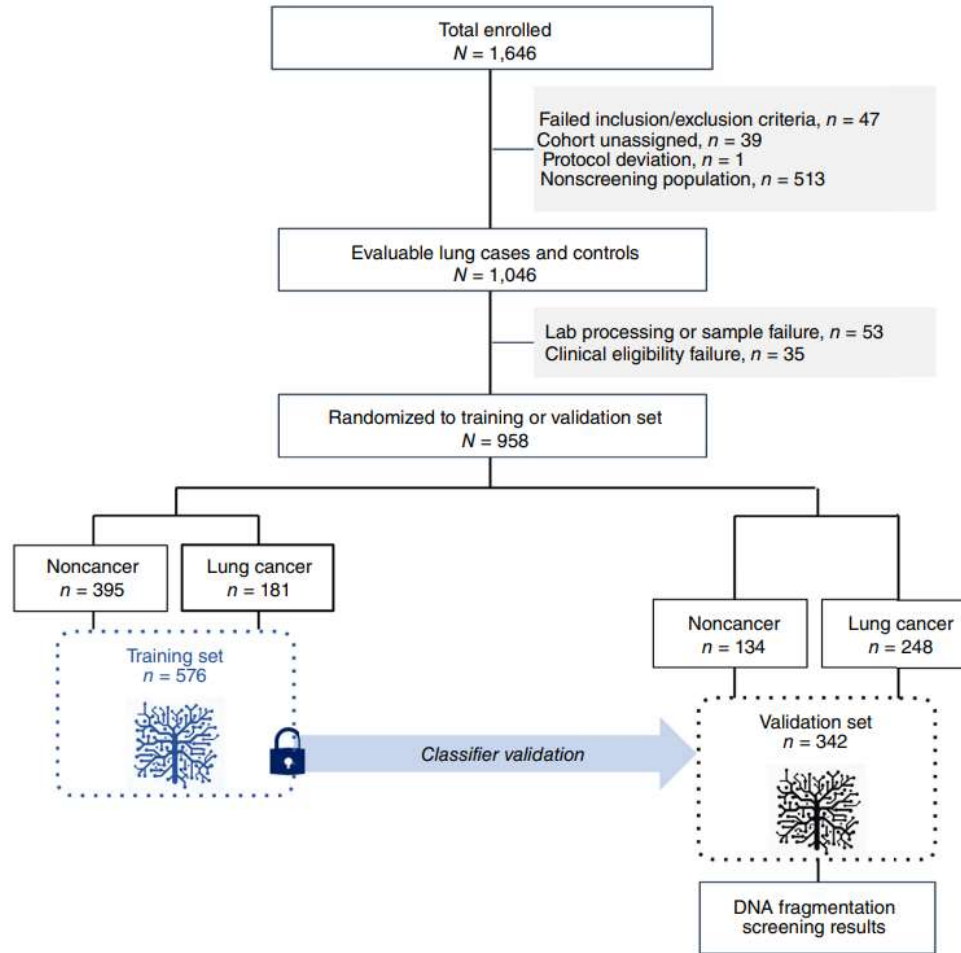


Mathios D, et al. *Nature Communications*, 2021;12(1):5060.



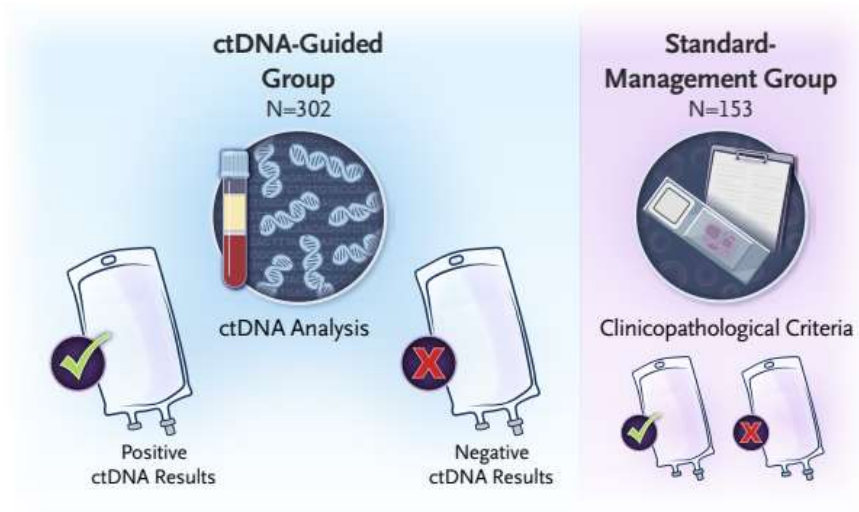
Nature. 2019 Jun;570(7761):385-389.

# FRAGMENTOMICS



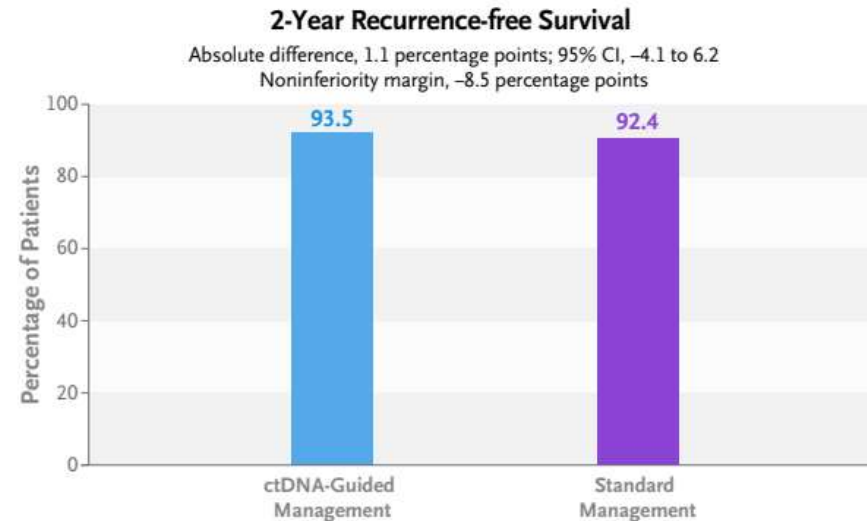
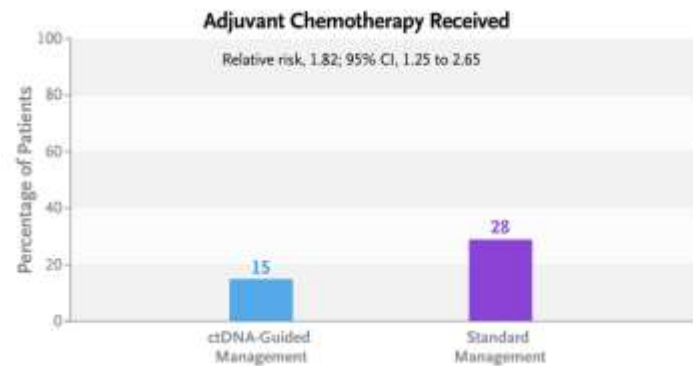
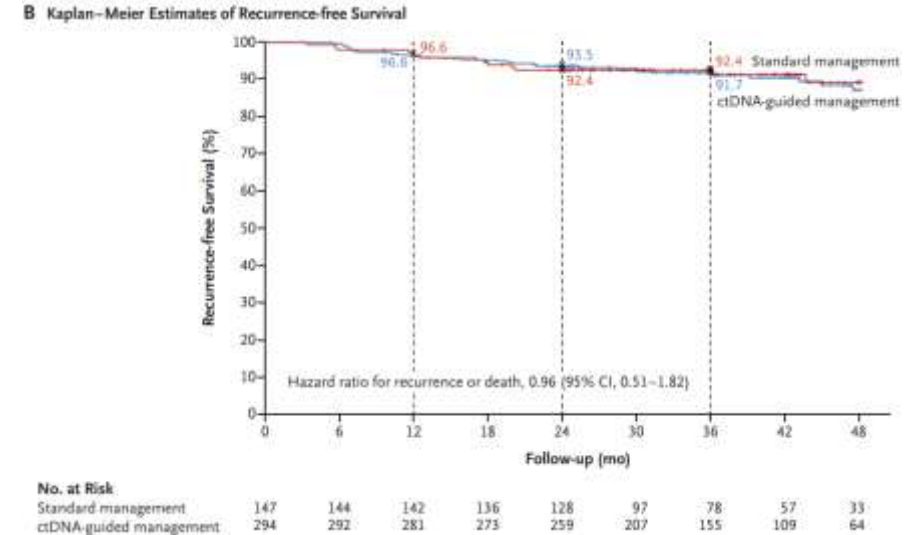
# Clinical Validation

<https://www.nejm.org/doi/full/10.1056/NEJMoa2200075#ap2>



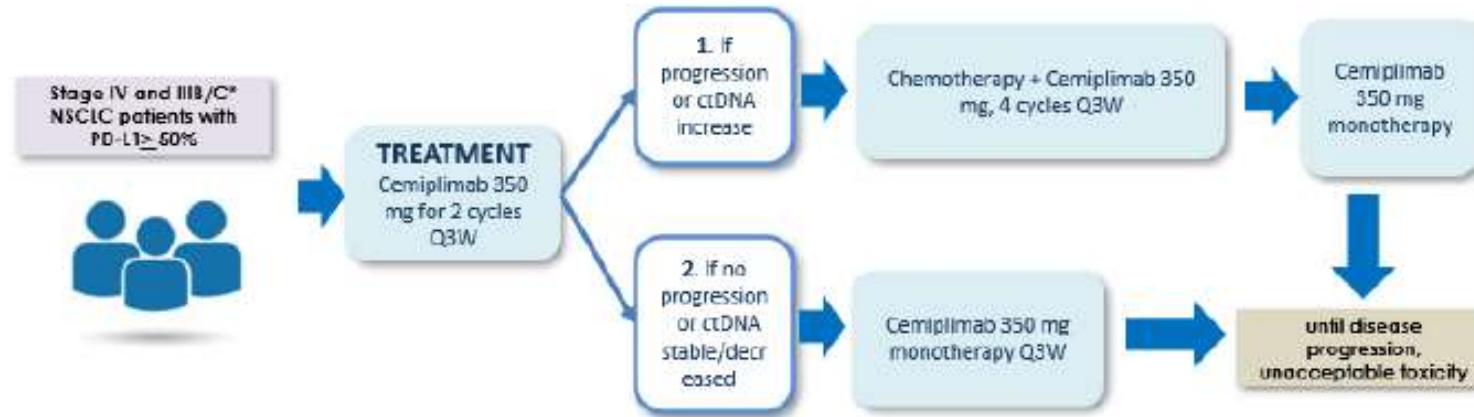
**Stage II Colon Cancer**

- R0 resection
- ECOG 0 – 2
- Staging CT within 8 weeks
- Provision of adequate tumor tissue within 4 weeks post-op
- No synchronous colorectal cancer



# Clinical Validation

## Palace



Primary endpoint: OS at 24 months in the ITT population



\*Stage IIIb/C not candidates for definitive Chemo/radiotherapy or surgical resection

26 pts

## External Quality Assessment (EQA) for Circulating free DNA (cfDNA) testing in Lung cancer 2023

*Table 5: Number of participating laboratories*

<b>Number of registrations</b>	357
<b>Number of withdrawals</b>	22
<b>Number of laboratories that did not submit results</b>	19*
<b>Total number participating laboratories</b>	316

\* Participation in this EQA is sponsored and has a limited number of available places; we request laboratories to only register if they are planning to submit results.

Sixteen laboratories using either the Oncomine™ Lung Cell-Free Total Nucleic Acid (cfTNA) Research Assay or the Oncomine™ Pan-Cancer Cell-Free Assay did not report the presence of the insertion in exon 20 of EGFR. There were other laboratories using these assays which did report the presence of the variant. Further investigation of this demonstrated that although this region of *EGFR* was included in the regions sequenced it was not in the “hotspot” file required to call variants using the kit bioinformatics. Many of these sixteen laboratories did not provide sufficient information in their reports to enable readers of the reports to determine that the assay used would not detect all variants in exon 20 of *EGFR* and therefore received deductions from their interpretation scores.

# Take home message

- Liquid biopsy sensitivity remains limited for MRD analysis.
- Specificity issues arise due to clonal hematopoiesis (CHIP).
- Tumor fraction estimation is critical and method-dependent.
- High pre-analytical and analytical variability; standardization is essential.
- Clinical validation is still limited.

16<sup>th</sup>  
CONGRESS  
*Lung* ON  
CANCER

BARCELONA  
27 / 28  
NOVEMBER 2025



# THANK YOU

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gecp  
lung cancer  
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